

Short communication

Z-isomers of 2-hydroxymethylcyclopropylidenemethyl adenine (synadenol) and guanine (synguanol) are active against ganciclovir- and foscarnet-resistant human cytomegalovirus UL97 mutants

Fausto Baldanti^{a,b}, Antonella Sarasini^a, John C. Drach^c, Jiri Zemlicka^d,
Giuseppe Gerna^{a,*}

^a Servizio di Virologia, IRCCS Policlinico S. Matteo, 27100 Pavia, Italy

^b Laboratori Sperimentali di Ricerca, IRCCS Policlinico S. Matteo, 27100 Pavia, Italy

^c University of Michigan, Ann Arbor, MI 48109-1078, USA

^d Karmanos Cancer Institute, Wayne State University, Detroit, MI 48201, USA

Received 7 May 2002; accepted 2 August 2002

Abstract

Emergence of drug-resistant human cytomegalovirus (HCMV) strains is a substantial problem during treatment of HCMV infections in immunocompromised patients. The Z-isomers of 2-hydroxymethylcyclopropylidenemethyl adenine (synadenol) and guanine (synguanol) were previously shown to be potent inhibitors of AD169 and Towne HCMV reference strains and postulated to share a common phosphorylation pathway with ganciclovir (GCV) possibly involving the UL97-encoded phosphotransferase. Analysis of synadenol and synguanol susceptibility of a series of HCMV isolates from immunocompromised untreated patients and from patients with treatment failure due to the emergence of GCV- and foscarnet (PFA)-resistant HCMV strains demonstrated that synadenol and synguanol are potent inhibitors of clinical HCMV isolates and are highly effective against both GCV- and PFA-resistant isolates. These results together with those showing resistance of a UL97 knock-out HCMV mutant to GCV as well as synadenol and synguanol suggest the involvement of UL97 phosphotransferase in synadenol and synguanol anabolism but with a substrate specificity different from that of GCV.

© 2002 Elsevier Science B.V. All rights reserved.

Keywords: Synadenol; Synguanol; Ganciclovir-resistance; Foscarnet-resistance; HCMV drug-susceptibility; HCMV UL97 phosphotransferase

Human cytomegalovirus (HCMV) is a major opportunistic pathogen in immunocompromised patients causing severe disseminated infections often associated with multiple organ localizations in untreated AIDS patients and in solid organ or

* Corresponding author. Tel.: +39-382-502644; fax: +39-382-502599

E-mail address: g.gerna@smatteo.pv.it (G. Gerna).

bone marrow transplant recipients (Jacobson and Mills, 1988). Antiviral drugs currently licensed for treatment of HCMV infections include ganciclovir (GCV), foscarnet (PFA) and cidofovir. However, the emergence of drug-resistant HCMV strains has been reported in AIDS patients (Baldanti et al., 1995, 1996, 1998a,c; Chou et al., 1995a,b, 1998, 2002; Drew et al., 1999; Erice et al., 1989; Gerna et al., 1992; Jabs et al., 2001; Smith et al., 1998) as well as transplant recipients (Baldanti et al., 1998b; Bienvenu et al., 2000; Eckle et al., 2000; Knox et al., 1991; Limaye et al., 2000; Lurain et al., 1996; Rosen et al., 1997). Indeed, after the introduction of highly active antiretroviral therapies (HAART), the emergence of HCMV drug-resistant strains has been more frequently reported in transplant recipients, while the emergence of drug-resistant HCMV strains in HAART-treated individuals is to date only anecdotal (Baldanti et al., 2002). Thus, development of new anti-HCMV compounds with different mechanisms of action is still warranted.

The Z-isomers of 2-hydroxymethylcyclopropylidenemethyl purines and pyrimidines were shown to be potent inhibitors of human and murine cytomegalovirus, Epstein–Barr virus and varicella zoster virus (Qiu et al., 1998; Rybak et al., 1999, 2000). In particular, synadenol and synguanol were active against HCMV reference strains Towne and AD169 with an IC_{50} value of 2 μ M for both compounds in a plaque reduction assay (Drach et al., 1997). Addition of the drugs to cells at different times post-infection indicated that these compounds act at a similar time point as GCV, thus suggesting a mechanism of action similar to that of GCV. In addition, *in vitro* phosphorylation of GCV was inhibited by synadenol and synguanol, suggesting a common anabolism pathway for the three compounds (Drach et al., 1997).

In this study, the synadenol and synguanol susceptibilities of a series of HCMV isolates from immunocompromised untreated individuals and a series of GCV-resistant and double (GCV and PFA)-resistant HCMV isolates from AIDS patients carrying mutations in the UL97-encoded viral phosphotransferase and in both the UL97-encoded phosphotransferase and the UL54-en-

coded viral DNA polymerase, respectively, are reported. In parallel, an *in vitro* generated UL97 knock-out mutant was also analyzed.

HCMV isolates VR5611, VR6189 and VR6200 were recovered from blood of a heart transplant recipient and two patients with AIDS, respectively, who did not receive any previous treatment for HCMV disease. GCV-resistant HCMV isolates VR3480, VR4760, VR4990, VR4991, VR5120, VR5406 and VR6264 were recovered from blood of patients with AIDS and HCMV disease showing virologic failure on GCV treatment. The reported series of GCV-resistant clinical isolates was selected for this study because each of them showed the presence of a different mutation in UL97 (Table 1). Each mutation was proven by marker transfer experiments to confer a high degree of GCV-resistance (Baldanti et al., 1995, 1996, 1998c; Chou et al., 1995a, 2002; Lurain et al., 1994). In addition, isolates VR4760 and VR5120 were also shown to be PFA-resistant because of a mutation (V715M) in the viral DNA polymerase (Baldanti et al., 1996). Finally, the HCMV reference laboratory strain AD169 and an AD169-derived UL97 knock-out mutant lacking GCV-phosphorylating activity (Prichard et al., 1999) were included in the study.

GCV, synadenol and synguanol susceptibilities of all HCMV isolates were determined in triplicate using a reported immediate-early plaque reduction assay (Gerna et al., 1992). Drug resistance was defined as an increase in ID_{50} values \geq threefold with respect to mean control values (Gerna et al., 1995).

Differences between synadenol and synguanol IC_{50} values of HCMV isolates from untreated patients and GCV-resistant isolates were analyzed using the *t*-test for unpaired data, and the relative potencies of the two analogs versus GCV-susceptible or GCV-resistant HCMV isolates were analyzed using the *t*-test for paired data.

Results of parallel GCV, synadenol and synguanol susceptibility testing of isolates from untreated patients or GCV-resistant isolates are reported in Table 1. In detail, synadenol and synguanol IC_{50} values for the three HCMV isolates from untreated patients were in the same range as GCV IC_{50} values ($P > 0.05$). Thus, these new analogs

Table 1

Synadenol and synguanol susceptibilities of HCMV isolates from untreated and GCV-treated immunocompromised patients

HCMV isolate ^a	UL97 mutation	Synadenol (μM) ^b		Synguanol (μM) ^b		GCV (μM) ^b	
		IC ₅₀	IC ₉₀	IC ₅₀	IC ₉₀	IC ₅₀	IC ₉₀
<i>GCV-susceptible</i>							
VR5611	None	1.7	5.0	1.9	4.3	2.9	6.0
VR6189	None	1.8	4.7	2.0	4.9	1.8	3.5
VR6200	None	1.6	4.3	2.0	4.8	3.3	4.1
Mean value ±SD		1.7 ±0.1	4.6 ±0.2	2.0 ±0.1	4.7 ±0.3	2.7 ±0.8	4.5 ±1.3
<i>GCV-resistant</i>							
VR4760 ^c	M460I	2.4	6.9	2.0	4.5	40.0	93.0
VR4991	C592G	1.7	4.9	1.1	4.7	25.0	92.5
VR6264	A594V	0.6	2.1	4.0	7.8	15.0	33.7
VR3480	L595del	4.3	8.4	1.8	6.8	22.0	75.0
VR5120 ^c	L595S	2	4.2	2.2	4.8	25.0	72.5
VR5406	G598S	1.6	4.3	3.3	4.9	20.0	75.0
VR4990	C607Y	3.9	7.2	1.5	2.6	23.0	33.7
Mean value ±SD		2.4 ±1.3	5.4 ±2.2	2.3 ±1.0	5.1 ±1.7	24.2 ±7.7	67.9 ±24.8

^a The three GCV-susceptible HCMV isolates originated from untreated immunocompromised patients (one heart transplant recipient and two patients with AIDS). The seven GCV-resistant HCMV isolates with different UL97 mutations were recovered from as many AIDS patients failing GCV therapy.

^b IC₅₀ and IC₉₀ represent the 50 and 90% inhibitory concentrations, respectively.

^c VR4760 and VR5120 were also shown to be PFA-resistant (IC₅₀ 498.0 and 520.0 μM , respectively), because of the V715M change in the viral DNA polymerase (Baldanti et al., 1996).

showed a potency in inhibiting replication of HCMV clinical isolates in cell culture that was comparable to that of GCV. In addition, comparable IC₉₀ values were found for the three drugs.

Moreover, as shown in Table 1, mean synadenol and synguanol IC₅₀ and IC₉₀ values for GCV-resistant HCMV isolates were not significantly different from those found for control HCMV isolates from untreated patients ($P > 0.05$). In addition, synadenol and synguanol showed a comparable potency in inhibiting replication of GCV-resistant isolates. Specifically, mean synadenol and synguanol IC₅₀ values were 2.4 ± 1.3 and 2.3 ± 1.0 ($P > 0.05$), and the corresponding IC₉₀ values were 5.4 ± 2.2 and 5.1 ± 1.7 ($P > 0.05$), respectively.

Table 2 shows the comparison among GCV, synadenol and synguanol susceptibilities of HCMV laboratory reference strain AD169 and an AD169-derived mutant (RCA97.19) with a large deletion in UL97 (Prichard et al., 1999).

Table 2

Activity of GCV, synadenol and synguanol on a HCMV laboratory reference strain and a UL97 knock-out mutant

HCMV strain ^a	IC ₅₀ (μM) ^b		
	GCV	Synadenol	Synguanol
AD169	3.8 \pm 2.0	1.2 \pm 0.4	2.4 \pm 1.0
RCA97.19	19.6 \pm 8.6	3.9 \pm 0.9	9.6 \pm 4.0
IC ₅₀ fold increase	5.2 \pm 0.6	3.2 \pm 0.7	4.0 \pm 0.0

^a HCMV AD169, laboratory reference strain; RCA97.19, AD169-derived UL97 knock-out mutant (kindly provided by Dr Mark Prichard, Aviron, Mountain View, CA).

^b 50% Inhibitory concentration; data represent mean values (\pm SD) of three different experiments.

The observed fivefold increase in GCV IC₅₀ value of RCA97.19 could account for a high level of GCV-resistance. In fact, as shown in Table 1, a

similar increase in GCV IC₅₀ levels with respect to mean GCV IC₅₀ control values was observed in clinical GCV-resistant isolates (mean = 8.9-fold, range = 5.5–14.8). This result is likely to be related to the reported lack of GCV phosphorylation in cells infected with RCA97.19 (Prichard et al., 1999). In parallel, a consistent increase in synadenol and synguanol IC₅₀ levels (3.2 and 4.0 fold increase, respectively) was observed. These results support the involvement of the HCMV UL97 phosphotransferase in synadenol and synguanol anabolism. In fact, previous in vitro phosphorylation studies suggested a common anabolism pathway for synadenol, synguanol and GCV on the basis of competition experiments (Drach et al., 1997). In addition, uninfected cells did not show synadenol and synguanol phosphorylation activity (Drach et al., 1997).

It is interesting that UL97 phosphotransferase appears to be able to phosphorylate nucleoside analogs other than GCV. In fact, recent studies have shown that UL97 does not appear to be a nucleoside kinase (He et al., 1997; Michel et al., 1996, 1998), but it does contain functional regions homologous to conserved domains characteristic of protein kinases (Michel et al., 1998, 1999). However, the role of UL97 phosphotransferase appears to be different in the phosphorylation process of synadenol and synguanol compared to GCV. In fact, our data demonstrate that UL97 residues involved in GCV recognition and processing do not exert the same function for the new analogs, as shown by the susceptibility of all GCV-resistant UL97 mutants to both compounds. In addition, we observed that UL97 deletion had a great impact (fivefold increase) in GCV resistance of RCA97.19 with respect to its parental strain AD169.

In conclusion, synadenol and synguanol were shown to be potent inhibitors of replication of HCMV clinical isolates. In addition, these analogs were found to display full activity against a panel of GCV-resistant HCMV isolates carrying different mutations in the UL97-encoded phosphotransferase. Finally, although sharing a UL97-mediated phosphorylation process, synadenol and synguanol show a substrate specificity different from that of GCV. From the data presented here, synadenol

and synguanol may seem promising for the treatment of GCV-resistant HCMV infections.

Acknowledgements

We thank Mark Prichard, Aviron, Mountain View, CA, for providing HCMV UL97 knock-out mutant RCA97.19 and Linda D'Arrigo for revision of the English. This work was partially supported by Ministero della Sanità, Istituto Superiore di Sanità, Programma Nazionale AIDS (grant no. 50D.12), Ricerca Finalizzata (grant no. 820RFM99/01), and by grant P01-AI46390 from N.I.H.

References

- Baldanti, F., Silini, E., Sarasini, A., Talarico, C.L., Stanat, S.C., Biron, K.K., Furione, M., Bono, F., Palù, G., Gerna, G., 1995. A three-nucleotide deletion in the UL97 open reading frame is responsible for the ganciclovir resistance of a human cytomegalovirus clinical isolate. *J. Virol.* 69, 796–800.
- Baldanti, F., Underwood, M.R., Stanat, S.C., Biron, K.K., Chou, S., Sarasini, A., Silini, E., Gerna, G., 1996. Single amino acid changes in the DNA polymerase confer foscarnet resistance and slow-growth phenotype, while mutations in the UL97-encoded phosphotransferase confer ganciclovir resistance in three double-resistant human cytomegalovirus strains recovered from patients with AIDS. *J. Virol.* 70, 1390–1395.
- Baldanti, F., Simoncini, L., Talarico, C.L., Sarasini, A., Biron, K.K., Gerna, G., 1998. Emergence of a ganciclovir-resistant human cytomegalovirus strain with a new UL97 mutation in an AIDS patient. *AIDS* 12, 816–818.
- Baldanti, F., Simoncini, L., Sarasini, A., Zavattoni, M., Grossi, P., Revello, M.G., Gerna, G., 1998. Ganciclovir resistance as a result of oral ganciclovir in a heart transplant recipient with multiple human cytomegalovirus strains in blood. *Transplantation* 66, 324–329.
- Baldanti, F., Underwood, M., Talarico, C.L., Simoncini, L., Sarasini, A., Biron, K.K., Gerna, G., 1998. The Cys607 → Tyr change in the UL97 phosphotransferase confers ganciclovir resistance to two human cytomegalovirus strains recovered from two immunocompromised patients. *Antimicrob. Agents Chemother.* 42, 444–446.
- Baldanti, F., Paolucci, S., Parisi, A., Meroni, L., Gerna, G., 2002. Emergence of multiple drug-resistant human cytomegalovirus variants in two patients with human immunodeficiency virus infection unresponsive to highly active antiretroviral therapy. *Clin. Infect. Dis.* 34, 1146–1149.

- Bienvenu, B., Thervet, E., Bedrossian, J., Scieux, C., Mazeron, M.C., Thouvenot, D., Legendre, C., 2000. Development of cytomegalovirus resistance to ganciclovir after oral maintenance treatment in a renal transplant recipient. *Transplantation* 69, 182–184.
- Chou, S., Erice, A., Jordan, M.C., Vercellotti, G.M., Michels, K.R., Talarico, C.L., Stanat, S.C., Biron, K.K., 1995. Analysis of the UL97 phosphotransferase coding sequence in clinical cytomegalovirus isolates and identification of mutations conferring ganciclovir resistance. *J. Infect. Dis.* 171, 576–583.
- Chou, S., Guentzel, S., Michels, K.R., Miner, R.C., Drew, W.L., 1995. Frequency of UL97 phosphotransferase mutations related to ganciclovir resistance in clinical cytomegalovirus isolates. *J. Infect. Dis.* 172, 239–242.
- Chou, S., Marousek, G., Parenti, D.M., Gordon, S.M., LaVoy, A.G., Ross, J.G., Miner, R.C., Drew, W.L., 1998. Mutation in region III of the DNA polymerase gene conferring foscarnet resistance in cytomegalovirus isolates from 3 subjects receiving prolonged antiviral therapy. *J. Infect. Dis.* 178, 526–530.
- Chou, S., Waldemer, R.H., Senters, A.E., Michels, K.S., Kemble, G.W., Miner, R.C., Drew, W.L., 2002. Cytomegalovirus UL97 phosphotransferase mutations that affect susceptibility to ganciclovir. *J. Infect. Dis.* 185, 162–169.
- Drach J.C., Fan, B.Y., Ptak, R.G., Breitenbach, J.M., Borysko, K.Z., Qiu, Y.-L., Zemlicka, J. 1997. Selective activity of 2-hydroxymethylcyclopropylidenemethyl purines against human cytomegalovirus. *Antiviral Res.* 34: A83 (10th International Conference on Antiviral Research, Atlanta, GA, USA, abstract 147).
- Drew, W.L., Stempien, M.J., Andrews, J., Shadman, A., Tan, S.J., Miner, R., Buhles, W., 1999. Cytomegalovirus (CMV) resistance in patients with CMV retinitis and AIDS treated with oral or intravenous ganciclovir. *J. Infect. Dis.* 179, 1352–1355.
- Eckle, T., Prix, L., Jahn, G., Klingebiel, T., Handgretinger, R., Selle, B., Hamprecht, K., 2000. Drug-resistant human cytomegalovirus infection in children after allogeneic stem cell transplantation may have different clinical outcomes. *Blood* 96, 3286–3289.
- Erice, A., Chou, S., Biron, K.K., Stanat, S.C., Balfour, H.H., Jordan, M.C., Jr., 1989. Progressive disease due to ganciclovir-resistant cytomegalovirus in immunocompromised patients. *N. Engl. J. Med.* 320, 289–293.
- Gerna, G., Baldanti, F., Zavattoni, M., Sarasini, A., Percivalle, E., Revello, M.G., 1992. Monitoring of ganciclovir sensitivity of multiple human cytomegalovirus strains coinfecting blood of an AIDS patient by an immediate-early antigen plaque assay. *Antiviral Res.* 19, 333–345.
- Gerna, G., Sarasini, A., Percivalle, E., Zavattoni, M., Baldanti, F., Revello, M.G., 1995. Rapid screening for resistance to ganciclovir and foscarnet of primary isolates of human cytomegalovirus from culture-positive blood samples. *J. Clin. Microbiol.* 33, 738–741.
- He, Z., He, Y., Kim, Y., Chu, L., Ohmsted, C., Biron, K.K., Coen, D.M., 1997. The human cytomegalovirus UL97 protein is a protein kinase that autophosphorylates on serines and threonines. *J. Virol.* 71, 405–411.
- Jabs, D.A., Martin, B.K., Forman, M.S., Dunn, J.P., Davis, J.L., Weinberg, D.V., Biron, K.K., Baldanti, F., 2001. Mutations conferring ganciclovir resistance in a cohort of patients with acquired immunodeficiency syndrome and cytomegalovirus retinitis. *J. Infect. Dis.* 183, 333–337.
- Jacobson, M.A., Mills, J., 1988. Serious cytomegalovirus disease in the acquired immunodeficiency syndrome (AIDS): clinical findings, diagnosis and treatment. *Ann. Intern. Med.* 108, 585–594.
- Knox, K.K., Drobyski, W.R., Carrigan, D.R., 1991. Cytomegalovirus isolate resistant to ganciclovir and foscarnet from a marrow transplant patient. *Lancet* 337, 1292–1293.
- Limaye, A.P., Corey, L., Koelle, D.M., Davis, C.L., Boeckh, M., 2000. Emergence of ganciclovir disease among recipients of solid-organ transplants. *Lancet* 356, 609–610.
- Lurain, N.S., Spafford, L.E., Thompson, K.D., 1994. Mutation in the UL97 open reading frame of human cytomegalovirus resistant to ganciclovir. *J. Virol.* 68, 4427–4431.
- Lurain, N.S., Ammons, H.C., Kapell, K.S., Yeldandi, V.V., Garritty, E.R., O'Keefe, J.P., 1996. Molecular analysis of human cytomegalovirus strains from two lung transplant recipients with the same donor. *Transplantation* 62, 497–502.
- Michel, D., Kramer, S., Höhn, S., Schaarschmidt, P., Wunderlich, K., Mertens, T., 1999. Amino acids of conserved kinase motifs of cytomegalovirus protein UL97 are essential for autophosphorylation. *J. Virol.* 73, 8898–8901.
- Michel, D., Pavic, I., Zimmermann, A., Haupt, E., Wunderlich, K., Heuschmid, M., Mertens, T., 1996. The UL97 gene product of human cytomegalovirus is an early-late protein with a nuclear localization but is not a nucleoside kinase. *J. Virol.* 70, 6340–6347.
- Michel, D., Schaarschmidt, P., Wunderlich, K., Heuschmid, M., Simoncini, L., Mühlberger, D., Zimmermann, A., Pavic, I., Mertens, T., 1998. Functional regions of the human cytomegalovirus protein pUL97 involved in nuclear localization and phosphorylation of ganciclovir and pUL97 itself. *J. Gen. Virol.* 79, 2105–2112.
- Prichard, M.N., Gao, N., Jairath, S., Mulamba, G., Krosky, P., Coen, D.M., Parker, B.O., Pari, G.S., 1999. A recombinant human cytomegalovirus with a large deletion in UL97 has a severe replication deficiency. *J. Virol.* 73, 5663–5670.
- Qiu, Y.L., Ksebat, M.B., Ptak, R.G., Fan, B.Y., Breitenbach, J.M., Lin, J.S., Cheng, Y.C., Kern, E.R., Drach, J.C., Zemlicka, J., 1998. (Z)- and (E)-2-((hydroxymethyl)cyclopropylidene) methyladenine and -guanine. New nucleoside analogues with broad-spectrum antiviral activity. *J. Med. Chem.* 41, 10–23.
- Rybak, R.J., Hartline, C.B., Qiu, Y.L., Zemlicka, J., Harden, E., Marshall, G., Sommadossi, J.P., Kern, E.R., 2000. In vitro activities of methylenecyclopropane analogues of nucleosides and their phosphoralaninate prodrugs against cytomegalovirus and other herpesvirus infections. *Antimicrob. Agents Chemother.* 44, 1506–1511.

- Rybak, R.J., Zemlicka, J., Qiu, Y.L., Hartline, C.B., Kern, E.R., 1999. Effective treatment of murine cytomegalovirus infections with methylenecyclopropane analogues of nucleosides. *Antiviral Res.* 43, 175–188.
- Rosen, H.R., Benner, K.G., Flora, K.D., Rabkin, J.M., Orloff, S.L., Olyaei, A., Chou, S., 1997. Development of ganciclovir resistance during treatment of primary cytomegalovirus infection after liver transplantation. *Transplantation* 63, 476–478.
- Smith, I.L., Taskintuna, I., Rahhal, F.M., Powell, H.C., Ai, E., Mueller, A.J., Spector, S.A., Freeman, W.R., 1998. Clinical failure of CMV retinitis with intravitreal cidofovir is associated with antiviral resistance. *Arch. Ophthalmol.* 116, 178–185.